

Longevity of organic pepper (*Capsicum annuum* L.) seeds

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Abstract

This study was carried out to determine seed longevity in organic and conventionally produced pepper seeds from four different pepper cultivars. Seeds were stored at 20 ± 2 °C with 7.5 ± 0.5 seed moisture over 48 months. Longevity of seeds were evaluated with Ki (initial seed quality), P50 (half-viability period), σ (standard deviation of distribution of seed deaths in time), and regression coefficient values. The highest longevity was observed in 'Corbaci' and 'Yagli' cultivars, while 'Surmeli' and 'K. Dolma' were found to have shorter longevity. P50 was 43.4 and 40.2 months for 'Corbaci' and 34.9 and 39.7 months for 'Yagli' organic and conventional cultivars, respectively, whereas it was about 21.4 and 23.7 months in 'K. Dolma' and 'Surmeli' cultivars. Similarly, the highest σ and regression coefficient values were observed for 'Corbaci' and the lowest for 'Surmeli' cultivars. Organic and conventional pepper seed longevity was not different in the same species. Regression coefficient values were 0.043 in organic and 0.046 in conventional seeds for 'Corbaci'. Very close values were found between the two production systems for the other cultivars too. Results indicate that organic seeds had similar longevity to conventional ones. The main differences originated from the cultivars, not from the production system.

Keywords: germination; organic farming; pepper; seed storage

Introduction

Maintaining seed quality after harvest is important to obtain successful stand establishment and transplant production. This particularly needs attention when left over seeds that were produced a year before need to be used in the following year. Accordingly, seed longevity is a key aspect that impacts the yield and quality of plants in cultivation by means of its influence on field emergence frequency, seedling growth and seed structure (Turner and Merritt, 2009; Righetti *et al.*, 2015). Various factors are recognised to affect seed quality during storage but the most profound ones are seed moisture and temperature (Rajjou and Debeaujon, 2008). In general, the lower the seed moisture and temperature, the longer seed longevity. As a rule of thumb Harrington (1972) stated that every 1% decrease in seed moisture and 6 °C decrease in temperature doubles the storage life of the seed. Furthermore, oxygen is known to be a major element to adjust many physiological

activities caused by respiration, which accompanies seed deterioration (Wang *et al.*, 2018). Temperature and relative humidity were described to be connected with oxygen in determining the seed longevity while in storage (Ventura *et al.*, 2008; Groot *et al.*, 2012; Xin *et al.*, 2014; Colville and Pritchard, 2019). It is known that seed longevity declines during storage, with the seed germination percentage in the end reducing or resulting in an increased rate of abnormal seedlings.

Some species seeds genetically store better than others. Pepper is considered to have seeds with short life span (Priestley, 1986). It is a warm climate summer crop and is produced intensively in sub-tropical regions where high relative humidity and temperature prevail, which are the basic causes of seed deterioration (Adebisi and Abdul-Rafiu, 2016) during storage. Loss of seed viability during storage is a primary cause of poor stand establishment and reflects the warm and humid ambient storage environment and limited availability of controlled environment storage facilities.

Long-term food production needs sustainable agricultural practices including organic production. The entire production process conforms to the rules of organic production. The main features of organic crop production are the removal of chemical crop protectants, the use of organic inputs instead of chemical fertilisers and the certification of the organic production system (Groot *et al.*, 2004). Hence the first step in organic farming is producing high quality organic seeds with the same quality standards as conventional farming (Groot and Raaijmakers, 2018). Quality organic seed production is crucial since preventive measures to keep plant healthy, i.e. using pesticides and herbicides, are not allowed and competition with weeds is fierce (Sripathy *et al.*, 2012).

There are numerous studies on longevity of conventionally-produced pepper seeds (Thanos *et al.*, 1989; Sundstrom, 1990; Sanchez *et al.*, 1993; Basay *et al.*, 2006; Demir and Ozcoban, 2007; Demir *et al.*, 2009; Caixeta *et al.*, 2014; Panayotov and Aladjadjian, 2014; Adam *et al.*, 2018; Verma *et al.*, 2018). Nevertheless, organic seed production systems are different than conventional ones and require specific measures for production (Anonymous, 2010). Thus, the longevity of organic and conventionally produced seeds may differ. The purpose of the study was to determine the longevity of organically produced pepper seeds and compare with conventionally produced ones.

Materials and Methods

Seed production in organic and conventional systems

Seed production was performed in 2015 in the organic and conventional parcels of Atatürk Horticultural Central Research Institute - Yalova / Turkey. 'Surmeli', 'Kandil Dolma', 'Yaglik' and 'Corbaci' open-pollinated pepper cultivars were used as plant materials. In the Yalova region with temperate climatic conditions, the average air temperatures for the vegetation period in 2015 (March-October) ranged between 5.5 °C and 30.9 °C. During the vegetation period, different fertilizers were used depending on plant growth periods and the number of plant nutrients in the soil. Gentasol (30% total organic matter, 5% N, 3% K₂O, 2% P₂O₅) and Ormin K (30% K₂O, %5 organic matter) organic fertilisers were used for organic production and 20:20:20 (NPK) and potassium sulphate (50% K₂O) compound fertilisers were used for conventional production.

According to the results of soil analysis, the soil structures of the production parcels have a clayey structure. The soil pH was determined to be between 7.73-7.79. Therefore, the structures of the parcels have slightly alkaline character. The amount of organic matter was 2.03% in the organic parcel and 3.51% in the conventional parcel.

Seeds of four different pepper cultivars were sown in the second week of March in 2015. The study was managed according to the randomised complete block design with 4 replicates and each replicate consisted of 150 plants. Seedlings were planted in organic and conventional parcels in the second week of May. Plant spacing was 0.8 m between rows and 0.4 × 0.5 m between plants with double rows. Weekly fertilisation was performed

depending on the plant growth and the number of plant nutrients in the soil. Gentasol 20 L per 1000 m² was applied to the organic parcel and 20:20:20 (N-P-K) 2 kg per 1000 m² of compound fertilizer was given by drip irrigation to the conventional parcel. Potassium fertilisation was done during the fruit maturation stage. Accordingly, Ormin K 2 kg per 1000 m² was used in the organic parcel and potassium sulfate 1.5 kg per 1000 m² was applied to the conventional parcel.

Fruits were harvested at maturity stage (60-65 days after anthesis) for each cultivar. The seeds were extracted and dried at 30 °C until the seed moisture content reduced to 7-8%. The moisture content of dried seeds was determined according to ISTA (2016). Seeds were kept in a fridge until use. The mean weight of 1000 seeds were determined after the drying process.

Seed storage

Organic and conventional seeds, dried to 7-8% moisture content, were vacuum sealed in laminated aluminium packets and stored for 48 months at 20±2 °C. The longevity of organic and conventional pepper seeds was evaluated by conducting germination percentage and mean germination time studies at the regular intervals of every 4 months during the period of 48 months.

The germination test was set with 4 replicates and each replicate had 50 seeds for each production system with four cultivars. Two pieces of Whatman No: 1 blotting paper were placed in 9 cm petri dishes. Then 4 ml of distilled water was added after the seeds were placed in the petri dishes (ISTA, 2016). The germination test was executed at 25 °C in the dark and the percentage germination was determined using daily counts for 14 days. The germination papers were moistened as needed. Normal seedlings were assessed at the end of the germination test.

Mean germination time (MGT, days) was calculated according to the formula below (Ellis and Roberts, 1980):

$$MGT = \sum n.t / \sum n$$

where n = number of newly germinated seeds (2 mm radicle emerged) at time t, t = days from planting, and $\sum n$ = final germination.

Seed longevity analysis

Normal germination of organic and conventionally grown pepper seeds during storage (12 samples) were subjected to probit analysis as described by Ellis and Roberts (1981), with the convention that the probit of 50% normal seedling = 0. The seed viability equation was used to quantify seed deterioration rate as follows:

$$V = K_i - p/\sigma$$

Where v is the probit percentage of normal seedlings after a period of p (month) of storage at given seed moisture and temperature. K_i is the intercept on the probit viability axis before storage and is an index of the initial seed quality (combination effects of genetics and environmental factors on quality) in probit unit. The loss of viability is normally distributed in time; therefore, it can be quantified in terms of P50, half viability period or σ standard deviation of seed deaths in time (i.e. the time required to lose 1 probit of viability) at certain seed moisture and storage temperature. After converting normal percentages to probit in y axis, regression analysis was conducted and regression formulae ($y=a-bx$) were developed for each cultivar and production system.

Statistical analysis

The laboratory tests in the study were established in accordance with the experimental randomised parcel design. The percentage values obtained from these tests were subjected to \sqrt{n} transformation. JMP 8.0 Statistical package program was used for analysis. Data were assessed with analysis of variance for the main effects, whereby the means of values were compared using the Duncan multiple range test and least significant difference test ($p=0.05$).

Results

Initial total seed germination of both organically and conventionally produced seeds were much higher than 91% for all cultivars. Especially ‘Corbaci’ and ‘Yaglik’ cultivars had values above 94% of total and 90% of normal. The difference between organic and conventional seeds was less than 3% in total and 4% in normal germination (Table 1). Thousand seed weights ranged between 6.41 and 6.90 g for the cultivars. Seed moisture contents of the cultivars varied between 7.6 and 8.0%.

Table 1. Changes in initial normal and total germination percentages, seed moisture content in storage and 1000 seed weight of organically and conventionally grown four pepper cultivars

Cultivar	Seed types	Normal Germ. (%)	Total Germ. (%)	Seed mc. (%)	1000 Seed weight (g)
‘Surmeli’	Organic	88a	92a	7,9	6,66
	Conv.	90a	92a	8,0	6,41
‘K. Dolma’	Organic	87a	92a	7,6	6,79
	Conv.	89a	91a	7,7	6,56
‘Yaglik’	Organic	90a	94a	7,7	6,65
	Conv.	94a	95a	7,7	6,73
‘Corbaci’	Organic	95a	98a	7,8	6,90
	Conv.	93a	95a	7,7	6,78

The seed survival curves of organic and conventionally grown pepper seeds for the four cultivars are presented in Figure 1. The survival curves (normal germination plotted against storage period over 48 months) showed the typical sigmoidal pattern. Seeds of ‘Surmeli’ and ‘K. Dolma’ lost germinability completely after 40 months, while death of ‘Yaglik’ and ‘Corbaci’ seeds occurred by 48 months. Seed germination losses in ‘Surmeli’ and ‘K. Dolma’ were much faster in earlier sampling times of the storage than those of the other two species. In all cultivars, organic and conventional seed germination loss showed a very similar trend. The difference in germination was significant in just a few samplings, but not in the majority. Probit analyses of the survival curves are presented in Figure 2. Following probit analyses, survival curves converted to straight lines.

Table 2. Probit model parameters of four pepper cultivars produced in organic and conventional systems obtained from Figure 2

Cultivar	Seed types	Model parameters				Regression coefficient (b)
		Ki	P ₅₀	R ²	σ	
‘Surmeli’	Organic	1,36	23,3	0,84	17,1	0.058
	Conv.	1,39	23,1	0,81	16,6	0.060
‘K. Dolma’	Organic	1,49	21,4	0,84	12,4	0.069
	Conv.	1,61	23,7	0,88	16,6	0.067
‘Yaglik’	Organic	1,60	34,9	0,74	21,6	0.045
	Conv.	1,66	39,7	0,85	23,9	0.041
‘Corbaci’	Organic	1,88	43,4	0,81	23,0	0.043
	Conv.	1,87	40,2	0,81	21,3	0.046

Ki: The intercept on the probit viability axis of the regression line, or an index of the initial seed quality **P₅₀:** Half viability period; **σ :** The days required to lose 1 probit of seed viability, **R²:** The coefficient of determination of the probit regression line

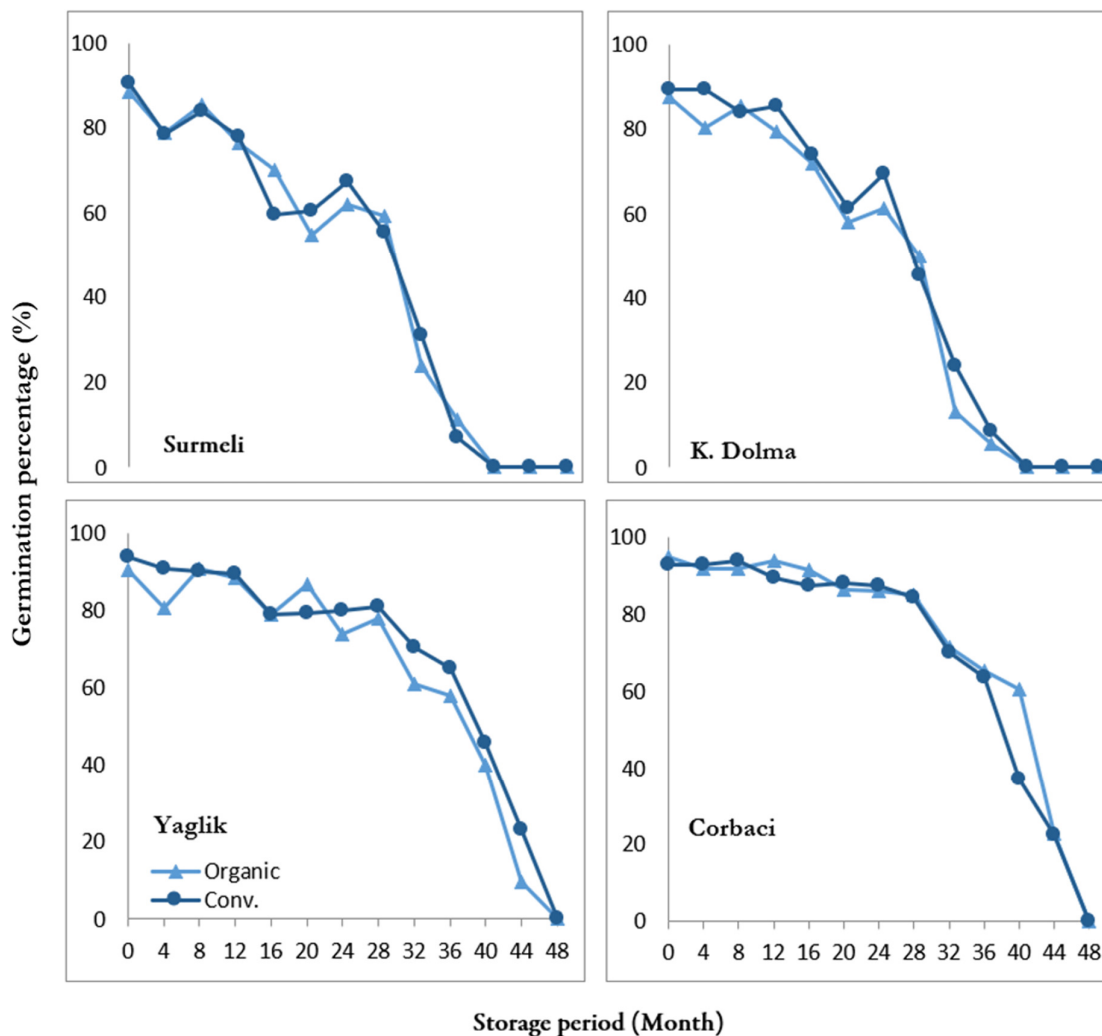


Figure 1. Survival curves of seeds of organically (Δ) and conventionally (●) grown pepper cultivars, 'Surmeli', 'K. Dolma', 'Yaglik' and 'Corbaci' stored at 20±2 °C with precise storage moisture contents

The greatest value of K_i was obtained from Corbaci as 1.87-1.88 in conventional and organic seeds, respectively (Table 2). The other cultivars were ranked as 'Yaglik', 'K. Dolma' and 'Surmeli' regarding K_i values. The minimum K_i value was observed in the Surmeli cultivar. Lower K_i values were related to shorter longevity as indicated by P_{50} and σ (Figure 4). There were differences among the cultivars but statistically there were no differences between organic and conventional seeds. 'Corbaci' also had the longest longevity as seen by P_{50} . It was followed by 'Yaglik' and 'Surmeli' which had the lowest values of 23.3 and 23.1 months in organic and conventional seeds. The σ values were larger (viability loss was slower) for 'Corbaci' and 'Yaglik' cultivars than for 'Surmeli' and 'K. Dolma'. There was no difference between σ values in relation to organic and conventional seeds (Table 2). Coefficient of regression (b) values also showed the rate of ageing was lowest in 'Corbaci' and 'Yaglik', but higher in 'Surmeli' and 'K. Dolma'.

Regarding seed vigour, mean germination time increased (Figures 3) as storage time extended. As the seeds were aged during storage, mean germination time increased from about 4-6 days to about 9-10 days. The difference between organic and conventional seeds was not significant in the majority of the samples. The greatest differences were seen in the 'Yaglik' cultivar in which organic seeds showed a more vigorous behaviour

during storage. Organic and conventional seed vigour was very similar in most of the samples, but the highest differences were seen in the 'Yaglik' cultivar.

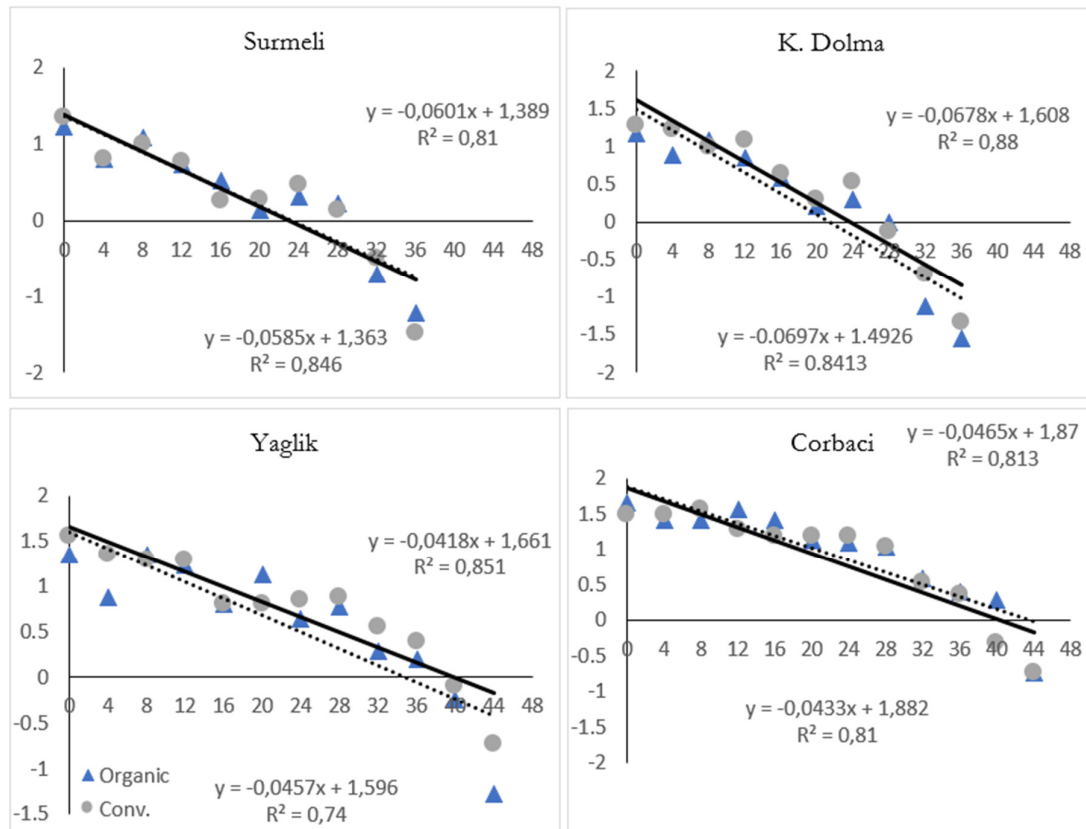


Figure 2. Probit values of germination percentages of survival curves in Figure 1. The estimates of K_i , P_{50} , σ and regression values were shown in Table 2

Discussion

The results of the study showed that the seed longevity of organic and conventional seeds was similar in four pepper cultivars. Differences were seen between the cultivars in terms of longevity. 'Surmeli' and 'K. Dolma' cultivars had faster seed deterioration compared to those of 'Yaglik' and 'Corbaci'. The longevity of any seed lot depends on various factors. One of which is the genetic structure (Wiebach *et al.*, 2019; Zhou *et al.*, 2019). Some species deteriorate faster than others. Pepper is also considered to be a faster deteriorating species (Passam and Lambropoulos, 1997). This is in overall agreement with comparisons of species concerning seed longevity by Priestley (1986). Ellis (1991) stated that some oily seeds such as lettuce and onion have shorter life-span than starchy ones such as peas, cereals and beans. But the conclusions of some reports contradict this finding. In our earlier work, watermelon seeds with higher oil content than pepper survived longer (Ozcoban and Demir, 2002). It can be stated that as the duration of seed storage rises, high amounts of peroxidation and oxidation of the lipids in the seed tends to reduce the volume of unsaturated fatty acids and soluble sugars (Li *et al.*, 2005). The rapid decline in seed viability in some cultivars or species compared to others requires special attention for some cultivar seeds. Incidence ageing is common in hot and humid environments particularly for pepper seeds that are grown in such regions.

Organic farming is gaining broad recognition as a system and the demand for organic products is constantly increasing due to perception by consumers as healthier and safer for the environment (Knight and Newman, 2013). Farmers are challenged by obtaining organic seeds with high quality. Consecutively, there has been significant progress in increasing the availability of organic seeds. Investments in organic seed research are rising (Anonymous, 2016). Therefore, the storage potential of organic seeds needs attention. Vegetables including peppers are grown by transplants and high-quality transplant production depends on the use of high-quality seeds (Groot *et al.*, 2005) because fast and uniform emergence is provided by use of highly vigorous seeds. The necessity to store seeds for use in the next production year may cause deterioration during the storage period, which is the main reason for low emergence and transplant size (Sano *et al.*, 2016). In our study, the gradual increase in MGT and reduction indicates that seeds lose emergence and transplant production potential (vigour) as storage extends. This can be an important aspect for production of organic transplant production (Rosso, 2005). High quality seeds are vital for successful transplant production in modules and crop establishment and weed competitiveness during the early stages of crop growth. Improved seed longevity in organic seeds is even more crucial (Groot *et al.*, 2004).

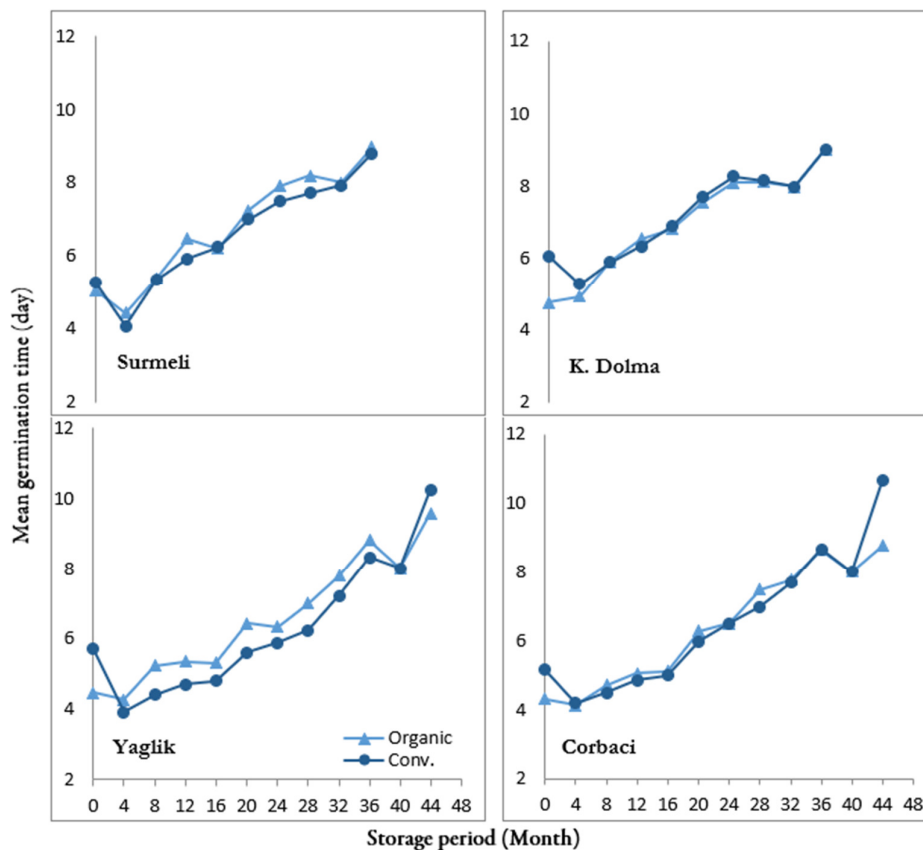


Figure 3. Changes in mean germination times of four pepper cultivars grown organically(Δ) and conventionally(●) systems during storage. Calculation was not included when seed germination was null

Mean germination times were extended in both seed production types. The MGT test is a well-known vigour test that is related to longevity of pepper seeds (Demir *et al.*, 2008). Seed vigour loss precedes germination loss during storage (Powell *et al.*, 1991). Our work also confirms this finding. Continuous loss of germination time and an increase in MGT during storage was seen in Figure 3. At the initial stages of sampling (4-12 months storage), germination levels in Yaglik and Corbaci were very high. But the time to germination and MGT values are increased gradually (Figure 3). This obviously shows that seed vigour changes precede

germination loss during storage, as seen in many different crop seeds (Abba and Lovato, 1999; Garcia *et al.*, 2006; Demirkaya *et al.*, 2010; Goyoaga *et al.*, 2011; Groot *et al.*, 2015; Demir *et al.*, 2016; Yin *et al.*, 2017; Zhou *et al.*, 2019).

There is a considerable variation in seed longevity among orthodox species which makes it possible to determine trends in relative seed longevity associated with climate, production systems or country of origin, for instance (Walters *et al.*, 2005; Probert *et al.*, 2007). There is much less variation within a species or even a genus. Some studies assumed that the rate of loss, i.e. standard deviation of distribution of seed death over time, does not differ between seed lots of the same species stored at the same moisture content and temperature (Ellis and Roberts, 1980). Thus, it was suggested that only the initial quality of a seed lot of Ki determines the longevity of a seed lot of a particular species in storage. Our results showed that Ki describes the seed quality and longevity potential of any seed lot that is produced by organic or conventional methods (Figure 4). It is an indicator of the combination effect of genotype and the pre-storage environment on seed quality, and in turn longevity (Rajjou *et al.*, 2008; Kochanek *et al.*, 2010; Zhou *et al.*, 2019). Both production systems had different Ki values but similar σ , p50, and regression coefficient values in similar storage conditions. This indicates that utmost attention should be paid to pre-storage seed production practices during organic and conventional seed production to obtain the maximum quality and longevity.

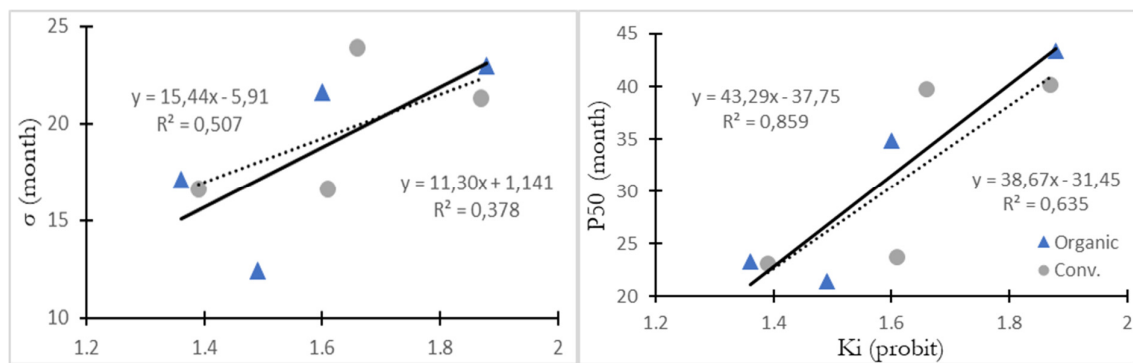


Figure 4. The relationship between Ki and σ and P50 values derived from Table 2

The four pepper cultivars used in this work have different fruit shapes. The one with shortest longevity of ‘Surmeli’ is a long green type and ‘K. Dolma’ is a bell shape. Both lose germinability faster during storage. Different seed lots even within the same genotype can differ substantially in longevity during experimental storage under similar conditions (same temperature and seed moisture) as a result of differences in the seed production environment conditions of harvest and the post-harvest drying and storage conditions (Ellis and Roberts, 1981). The seed lots were produced under either organic or conventional conditions and dried under identical conditions. The other difference can originate from maturation stages. Even though we harvested fruits when they were in the bright red mature stage, some cultivars may need longer maturation time on the mother plant to obtain maximum longevity (Zanakis *et al.*, 1993).

Conclusions

In conclusion, this study indicates that organic pepper seeds had the same seed longevity as conventionally grown ones. The basic differences in longevity originated from cultivar differences and genetic backgrounds. Accordingly, in spite of the fact that all the cultivation processes were the same within the production systems, the responses of the cultivars to longevity can still be different. Pre-storage factors and genetic structure should be taken into account when considering pepper seed longevity.

Authors' Contributions

KCY and ID were planned the experiments, discussed and interpreted the obtained results, like wise, KCY was designed manuscript according to writing style for the journal, EO was statistically analyzed the data and created the figures, ZG was controlled the manuscript in terms of grammatically and modal/formal convenience, errors, etc. All authors read and approved the final manuscript.

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Conflict of Interests

The authors declare that there are no conflicts of interest related to this article.

References

- Abba EJ, Lovato A (1999). Effect of seed storage temperature and relative humidity on maize (*Zea mays* L.) seed viability and vigour. *Seed Science and Technology* 27:101-114. <https://doi.org/10.1088/1755-1315/484/1/012116>
- Adam O, Abisoye O, Tolani U, Julius O, Ayooluwa O, Taiwo A (2018). Germination performance and vigour of pepper seeds stored in different environmental conditions at different storage periods. *Asian Journal of Biology* 5(2):1-6. <https://doi.org/10.9734/AJOB/2018/39791>
- Adebisi MA, Abdul-Rafiu AM (2016). Storage life of cayenne pepper (*Capsicum frutescens*) seeds following three drying methods. *Seed Technology* 37(1):33-42. <https://www.jstor.org/stable/26625370?seq=1>
- Anonymous (2010). Regulation on principles and application of organic agriculture. 18.08.2010 dated and 54 27676 numbered Official Gazette. <http://www.resmigazete.gov.tr/eskiler/2010/08/20100818-4.htm>.
- Anonymous (2016). State of organic seed. Retrieved March 25 2019 from <http://stateoforganicseed.org/wp-content/uploads/2017/01/SOS-2016-report-FINAL-DIGITAL.pdf>
- Basay S, Surmeli N, Okcu G, Demir I (2006). Changes in germination percentages, protein and lipid contents of primed pepper seeds during storage. *Acta Agriculturae Scandinavica Section B-Soil and Plant Science* 56(2):138-142. <https://doi.org/10.1080/09064710510029231>
- Caixeta F, von Pinho VRÉ, Guimarães RM, Pereira PHAR, Catão HCRM (2014). Physiological and biochemical alterations during germination and storage of habanero pepper seeds. *African Journal of Agricultural Research* 9(6):627-635. <https://doi.org/10.5897/AJAR2013.7133>
- Colville L, Pritchard HW (2019). Seed life span and food security. *New Phytologist* 224(2):557-562. <https://doi.org/10.1111/nph.16006>
- Demir I, Ozcoban M (2007). Dry and ultra-dry storage of pepper, aubergine, winter squash, summer squash, bean, cowpea, okra, onion, leek, cabbage, radish, lettuce and melon seeds at -20 °C and 20 °C over five years. *Seed Science and Technology* 35:165-175. <https://doi.org/10.15258/sst.2007.35.1.15>
- Demir I, Ermiş S, Mavi K, Matthews S (2008). Mean germination time of pepper seed lots (*Capsicum annuum* L.) predicts size and uniformity of seedlings in germination tests and transplant modules. *Seed Science and Technology* 36:21-30. <https://doi.org/10.15258/sst.2008.36.1.02>
- Demir I, Kenanoglu BB, Mavi K, Celikkol T, Hay F, Sariyildiz Z (2009). Derivation of constants (K_E , C_W) for the viability equation for pepper seeds and the subsequent test of its applicability. *HortScience* 44:1679-1682. <https://doi.org/10.21273/HORTSCI.44.6.1679>

- Demir I, Kara F, Ozden E, Hassanzadeh M (2016). The effects of seed moisture content and regional storage temperature on the longevity of two onion cultivars. VII International Symposium on Edible Alliaceae, 21-25 May, ActaHorticulturae, Niğde, Turkey pp 1143-1148. <https://doi.org/10.17660/ActaHortic.2016.1143.48>
- Demirkaya M, Dietz K J, Sivritepe HÖ (2010). Changes in antioxidant enzymes during ageing of onion seeds. Notulae Botanicae Horti Agrobotanici Cluj-Napoca 38(1):49-52. <https://doi.org/10.15835/nbha3814575>
- Ellis RH (1991). Longevity of seeds. HortScience 26(9):1119-1123.
- Ellis RH, Roberts EH (1980). Towards a rational basis for seed testing in seed production. Ed. P.D. Hebblethwaite, Butterworths, London, pp 605-635. <https://agris.fao.org/agris-search/search.do?recordID=US201301358784>
- Ellis RH, Roberts EH (1981). The quantification of aging and survival in orthodox seeds. Seed Science and Technology 9:373-409. <https://agris.fao.org/agris-search/search.do?recordID=XE8182678>
- Garcia I S, Souza A, Barbedo CJ, Dietrich SM, FigueiredoRCL (2006). Changes in soluble carbohydrates during storage of *Caesalpinia echinata* LAM. (Brazilwood) seeds, an endangered leguminous tree from the Brazilian Atlantic forest. Brazilian Journal of Biology 66:739-745. [10.1590/s1519-69842006000400018](https://doi.org/10.1590/s1519-69842006000400018)
- Goyoaga C, Burbano C, Cuadrado C (2011). Content and distribution of protein, sugars and inositol phosphates during the germination and seedling growth of two cultivars of *Vicia faba*. Journal of Food Composition and Analysis 24(3):391-397. <https://doi.org/10.1016/j.jfca.2010.11.002>
- Groot SPC, van der Wolf JWM, Jalink H, Langerak CJ, van den Bulk RW (2004). Challenges for the production of high quality organic seeds. Seed Testing International 127:12-15. <https://edepot.wur.nl/40952>
- Groot SPC, van den Bulk RW, Joost van der Burg W, Jalink H, Langerak CJ, van der Wolf JM (2005). Production of organic seeds: status, challenges and prospects. Seed Info, Official Newsletter of WANA Seed Network 9-12. <https://www.seedtest.org/upload/cms/user/ChallengesforOrganicSeed.pdf>
- Groot SPC, Surki AA, De VRCH, Kodde J (2012). Seed storage at elevated partial pressure of oxygen, a fast method for analysing seed ageing under dry conditions. Annals of Botany 110:1149-1159. <https://doi.org/10.1093/aob/mcs198>
- Groot SPC, de Groot L, Kodde J, van Treuren R (2015). Prolonging the longevity of ex situ conserved seeds by storage under anoxia. Plant Genetic Resources: Characterization and Utilization 13(1):18-26. <https://doi.org/10.1017/S1479262114000586>
- Groot SPC, Raaijmakers MHJE (2018). Organic seed production, certification and availability: Corp breeding and cultivation. In: Köpke U (Ed.), Improving organic crop cultivation Cambridge, UK. Burleigh Dodds Science Publishing Limited. <https://research.wur.nl/en/publications/organic-seed-production-certification-and-availability-corp-breed>
- Harrington JF (1972). Seed storage and longevity. Seed Biology 3:145-245.
- ISTA (2016). International rules for seed testing. Edition 2016. International Seed Testing Association, Bassersdorf, Switzerland. <https://www.seedtest.org/upload/cms/user/OGM15-05-Proposed-Changes-to-the-ISTA-Rules-for-2016.pdf>
- Kochanek J, Buckley YM, Probert RJ, Adkins SW, Steadman KJ (2010). Pre-zygotic parental environment modulates seed longevity. Austral Ecology 35:837-848. <https://doi.org/10.1111/j.1442-9993.2010.02118.x>
- Knight KW, Newman S (2013). Organic agriculture as environmental reform: a cross-national investigation. Society & Natural Resources 26(4):369-385. <https://doi.org/10.1080/08941920.2012.687070>
- Li X, Zou X, Liu Z (2005). On physiological and biochemical changes of artificially aged pepper seeds. Journal of Hunan Agricultural University 31(3):265-268. <https://europepmc.org/article/cba/599480>
- Ozcoban M, Demir I (2002). Longevity of pepper (*Capsicum annuum*) and watermelon (*Citrullus lanatus*) seeds in relation to seed moisture and storage temperature. Indian Journal of Agricultural Science 72:589-593.
- Panayotov N, Aladjadjian A (2014). Effect of long-term storage of pepper (*Capsicum annuum* L.) seeds on their viability measured by selected thermodynamic parameters. Acta Science Polonorum, Hortorum Cultus 13(2):151-162.
- Passam HC, Lambropoulos, EM (1997). Pepper seed longevity following production under high ambient temperature. Seed Science and Technology 25:177-185.
- Powell AA, Thornton JM, Mitchell JA (1991). Vigour differences in brassica seed and their significance to emergence and seedling variability. Journal of Agricultural Science 116:369-373. <https://doi.org/10.1017/S0021859600078187>
- Priestley DA (1986). Seed ageing. Cornell University Press, New York, NY. <https://doi.org/10.1017/S001447970001704X>

- Probert R, Adams J, Coneybeer J, Crawford A, Hay F (2007). Seed quality for conservation is critically affected by pre-storage factors. *Australian Journal of Botany* 55:326-335.
- Rajjou L, Debeaujon I (2008). Seed longevity: survival and maintenance of high germination ability of dry seeds. *Comptes Rendus Biologies* 331:796-805. <https://doi.org/10.1016/j.crv.2008.07.021>
- Rajjou L, Lovigny Y, Groot SPC, Belghazi M, Job C, Job D (2008). Proteome-wide characterization of seed aging in Arabidopsis: a comparison between artificial and natural aging protocols. *Plant Physiology* 148:620-641. <https://doi.org/10.1104/pp.108.123141>
- Righetti K, Vu JL, Pelletier S, Vu BL, Glaab E, Lalanne D, Buitink J (2015). Inference of longevity-related genes from a robust coexpression network of seed maturation identifies regulators linking seed storability to biotic defense-related pathways. *Plant Cell* 27:2692-2708. <https://doi.org/10.1105/tpc.15.00632>
- Rosso VM (2005). Organic vegetable transplant production. *HortScience* 40(3):623-628. <https://doi.org/10.21273/HORTSCI.40.6.1594A>
- Sanchez VM, Sundstrom FJ, McClure GN, Lang NS (1993). Fruit maturity, storage and postharvest maturation treatments affect bellpepper (*Capsicum annuum* L.) seed quality. *Scientia Horticulturae* 54(3):191-201. <https://agris.fao.org/agris-search/search.do?recordID=NL19930106355>
- Sano N, Rajjou L, North HM, Debeaujon I, Marionpoll A, Seo M (2016) Staying alive: Molecular aspects of seed longevity. *Plant and Cell Physiology* 57:660-674. <https://doi.org/10.1093/pcp/pcv186>
- Sripathy KV, Hosamani J, Bellundagi A, Prabhakar I (2012). Organic seed production. *Environment & Ecology* 30(1):102-105.
- Sundstrom FJ (1990). Seed moisture influence on tobasco pepper seed viability, vigour and dormancy during storage. *Seed Science and Technology* 18:179-185. <https://www.cabdirect.org/cabdirect/abstract/19920314172>
- Thanos CA, Georgiou K, Passam HC (1989). Osmoconditioning and ageing of pepper seeds during storage. *Oxford Journals Science and Mathematics Annals of Botany* 63:65-70.
- Turner S, Merritt D (2009). Seed germination and dormancy. *Plant Cell* 9:1055-1066. <https://doi.org/10.1105/tpc.9.7.1055>
- Ventura L, Donà M, Macovei A, Carbonera D, Buttafava A, Mondoni A, Balestrazzi A (2008). Understanding the mechanisms and kinetics of seed aging. *Plant Physiology and Biochemistry* 60:196-206. <https://doi.org/10.1093/conphys/cov026>
- Verma VK, Jha AK, Patel RK, Ngachan SV (2018). Studies on storage life, and effect of temperature and pre-sowing seed treatments on germination behaviour and maturity indices in King-chilli (*Capsicum* spp). *Indian Journal of Agricultural Sciences* 88 (8):1162-1167. <https://krishi.icar.gov.in/jsui/handle/123456789/14919>
- Walters C, Hill LM, Wheeler LJ (2005). Dying while dry: Kinetics and mechanisms of deterioration in desiccated organisms. *Integrative and Comparative Biology* 45:751-758. <https://doi.org/10.1093/icb/45.5.751>
- Wang W, He A, Peng S, Huang J, Cui K, Nie L (2018). The effect of storage condition and duration on the deterioration of primed rice seeds. *Frontiers in Plant Science* 9:172. <https://doi.org/10.3389/fpls.2018.00172>
- Wiebach J, Nagel M, Börner A, Altmann T, Riewe D (2019). Age-dependent loss of seed viability is associated with increased lipid oxidation and hydrolysis. *Plant Cell & Environment* 43(2):303-314. <https://doi.org/10.1111/pce.13651>
- Xin X, Tian Q, Yin G, Chen X, Zhang J, Ng S, Lu X (2014). Reduced mitochondrial and ascorbateglutathione activity after artificial ageing in soybean seed. *Journal of Plant Physiology* 171:140-147. <https://doi.org/10.1016/j.jplph.2013.09.016>
- Yin G, Xin X, Fu S, An M, Wu S (...), Lu X (2017). Proteomic and carbonylation profile analysis at the critical node of seed ageing in *Oryza sativa*. *Scientific Reports* 7:40611. <https://doi.org/10.1038/srep40611>
- Zanakis GN, Ellis RH, Summerfield RJ (1993). Response of seed longevity to moisture content in three genotypes of soyabean (*Glycine max*). *Experimental Agriculture* 29(4):449-459. <https://doi.org/10.1017/S0014479700021165>
- Zhou W, Chen F, Zhao S, Yang C, Meng Y, Shuai H (...), Shu K (2019). DA-6 promotes germination and seedling establishment from aged soybean seeds by mediating fatty acid metabolism and glycometabolism. *Journal of Experimental Botany* 70:101-114. <https://doi.org/10.1093/jxb/ery247>



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